Cell-ID 20-Plex Pd Barcoding Kit

Ask bigger questions.
Get higher-confidence answers.

Mass cytometry has opened new doors to discovery by giving researchers the power to interrogate more than 40 parameters simultaneously on millions of cells, unveiling new cell types and functions. With the launch of Fluidigm’s Cell-ID™ 20-Plex Pd Barcoding Kit (No. 201060), scientists can now vastly expand their throughput and get more robust data to reveal subtle phenotypes. The Cell-ID Kit enables barcoding of up to 20 samples, which are then stained and acquired as a single multiplexed sample (Figure 1). Perform your experiments with all the samples, time points and conditions you need to enable comprehensive cellular profiling.

Advantages of Barcoding

- **Throughput**: Reduces processing time throughout the entire experimental workflow.
- **Data quality**: Eliminates sample-to-sample variation introduced when samples are stained and collected individually. Discriminates cross-sample doublets by filtering out events with more than three Pd isotopes.
- **Sample conservation**: Enables efficient usage of precious samples by allowing you to combine multiple samples in one tube.
- **Accessibility**: Utilizes isotopes that are compatible with existing panel designs and can be used on all CyTOF systems.

“For all practical purposes, before CyTOF and mass-tag barcoding, the experiments that I wanted to do were impossible.”

—Dr. Eli Zunder, stem cell biologist, Stanford University.

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Figure 1. Each barcode consists of a unique combination of three palladium (Pd) isotopes.
Workflow

1. Barcode
2. Combine
3. Stain
4. Acquire
5. Debarcode
6. Analyze

Contents

<table>
<thead>
<tr>
<th>Component</th>
<th>Volume</th>
</tr>
</thead>
<tbody>
<tr>
<td>3 sets of 20 barcodes in PCR tubes</td>
<td>10 µL each tube</td>
</tr>
<tr>
<td>Maxpar® 5X Fix I Buffer</td>
<td>15 mL</td>
</tr>
<tr>
<td>Maxpar Cell Staining Buffer</td>
<td>500 mL</td>
</tr>
<tr>
<td>Maxpar 10X Barcode Perm Buffer</td>
<td>50 mL</td>
</tr>
<tr>
<td>Maxpar PBS</td>
<td>500 mL</td>
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</tbody>
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References
